



# The 28<sup>th</sup> Congress of the International Society for Forensic Genetics



PRAGUE, 9 – 13<sup>TH</sup> SEPTEMBER 2019, THE CZECH REPUBLIC, PRAGUE CONGRESS CENTRE

## Exploring DNA Interpretation Software Using the PROVEDIt Dataset

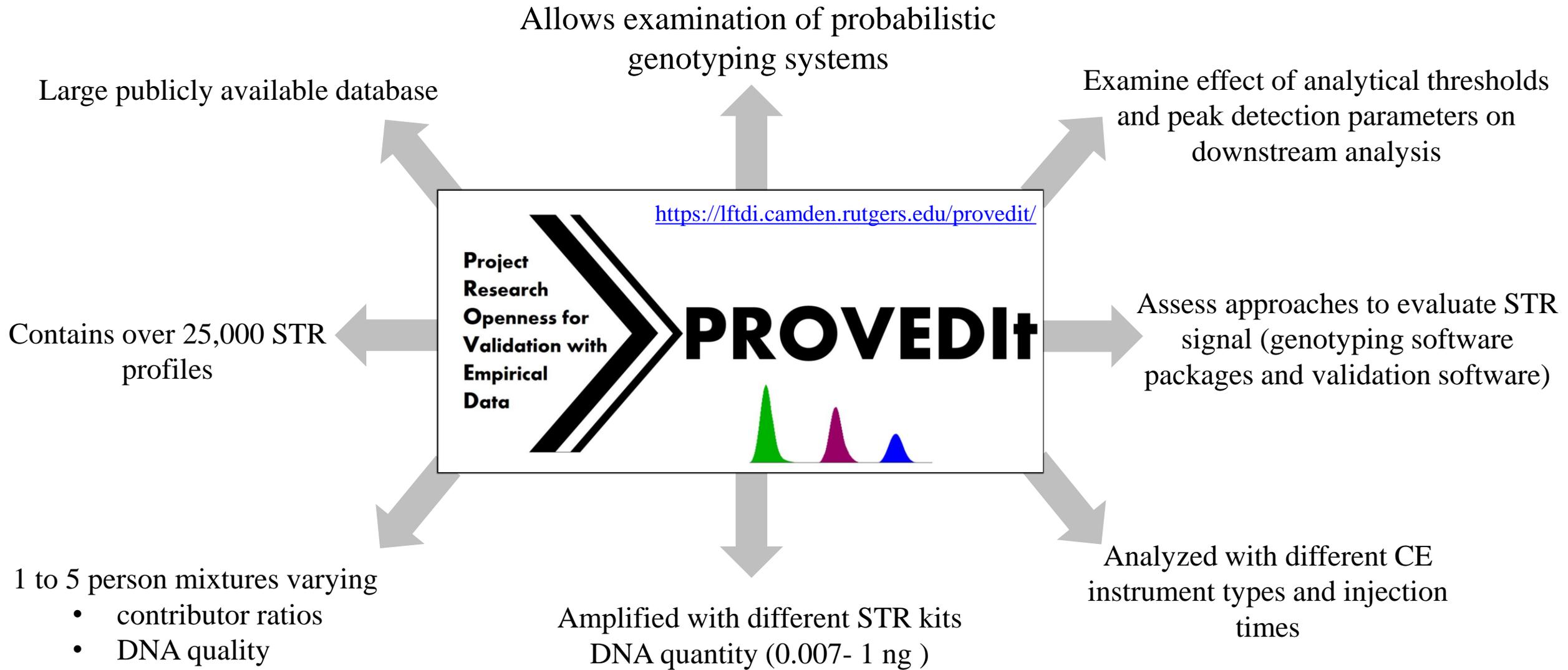
**Sarah Riman<sup>1</sup>; Hari Iyer<sup>2</sup>; Peter M. Vallone<sup>1</sup>**

<sup>1</sup>Applied Genetics Group, National Institute of Standards and Technology

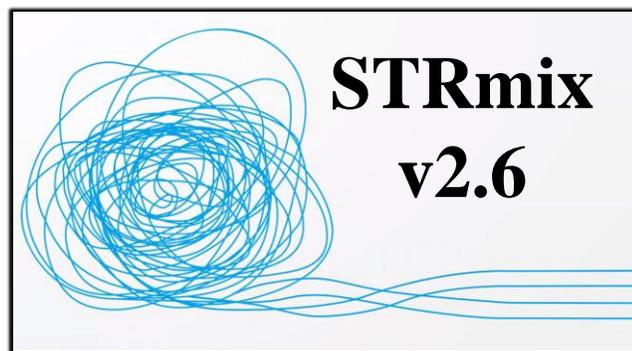
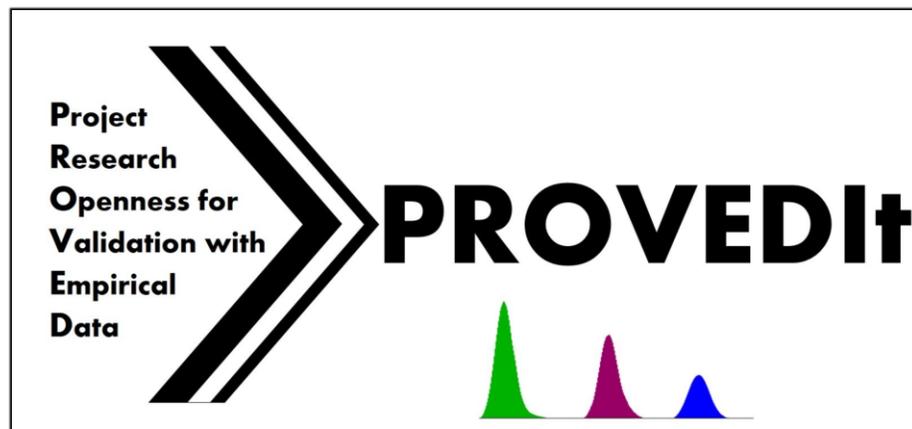
<sup>2</sup>Statistical Design, Analysis, and Modeling Group, National Institute of Standards and Technology



# Overview of PROVEDIt database



# Objective of this study



**EuroForMix v2.1.0**

*An open-source software for statistical DNA interpretation*

# Interpretation parameters used for each software

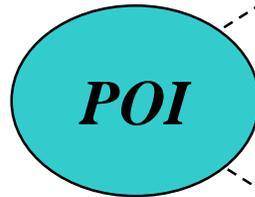
Software	
STRmix v2.6	<ul style="list-style-type: none"> <li>▪ <i>N-1, N-2 and N+1</i> stutter peaks were modeled</li> <li>▪ <i>Drop-in frequency</i> = 0.0015 and maximum cap = 180 RFU</li> <li>▪ <i>Saturation threshold</i> = 30,000 RFU</li> <li>▪ <i>MCMC settings</i>: 8 chains of 100,000 burn-in accepts, 50,000 post burn-in accepts per chain</li> <li>▪ Allelic, stutter, and locus-specific amplification efficiency variance were determined using <i>Model Maker</i> where over <i>300 single source profiles</i> of varying quality and quantity were assessed</li> <li>▪ The <a href="#">sub-source LR</a> is reported</li> </ul>
EuroForMix v2.1.0	<ul style="list-style-type: none"> <li>▪ <i>MLE</i> (Maximum likelihood estimation) approach</li> <li>▪ <i>Degradation and stutter models</i> jointly turned on</li> <li>▪ Default parameters, except for a 35 RFU <i>detection threshold</i>, <math>Pr(C) = 0.0015</math> and <math>\lambda = 0.018</math>.</li> <li>▪ The <a href="#">MLE based method LR</a> is reported</li> </ul>
Both software	<ul style="list-style-type: none"> <li>▪ Profiles were analyzed using the <i>per dye ATs</i></li> <li>▪ <i>NIST 1036-Caucasian</i> allele frequencies</li> <li>▪ <math>\theta</math> correction was applied using an <math>F_{st}(\theta) = 0.01</math></li> <li>▪ True <i>NOC</i> and <i>same propositions</i> were used in both software</li> </ul>

# Dataset used in our study

Kit (PCR cycle no.)			CE instrument (injection time)				
<b>GlobalFiler (29 cycles)</b>			<b>3500 (15 s)</b>				
Number of contributors	Mixture ratios	Pristine DNA	Degraded DNase I	Degraded Sonication	Damaged UV	Inhibited <u>Humic Acid</u>	Minor Contributor DNA amount (pg)
<b>2P</b> (16 unique individuals)	1:1	x	x	x	x	x	15; 30; 62; 125
	1:2	x	x				15; 30; 62; 125
	1:4	x	x	x	x	x	15; 30; 62; 125
	1:9	x	x	x	x	x	15; 30; 54; 62; 75
<b>Sum</b>		88	228	44	104	108	572
<b>3P</b> (21 unique individuals)	1:1:1	x	x	x	x	x	15; 30; 62; 125
	1:2:1	x	x				15; 30; 62; 125
	1:2:2	x	x				15; 30; 62; 125
	1:4:1	x	x	x	x	x	15; 30; 62; 125
	1:4:4	x	x	x	x	x	15; 30; 62; 83
	1:9:1	x	x				15; 30; 45; 62
	1:9:9	x	x				15; 26; 30; 40
<b>Sum</b>		114	324	72	138	162	810

# Dataset used in our study

Kit (PCR cycle no.)	CE instrument (injection time)
<b>GlobalFiler (29 cycles)</b>	<b>3500 (15 s)</b>
<b>Number of contributors</b>	
<b>2P</b>	
$H_p = \text{POI} + U_1$ $H_d = U_1 + U_2$	
<b>Sum</b>	
<b>3P</b>	
$H_p = \text{POI} + U_1 + U_2$ $H_d = U_1 + U_2 + U_3$	
<b>Sum</b>	



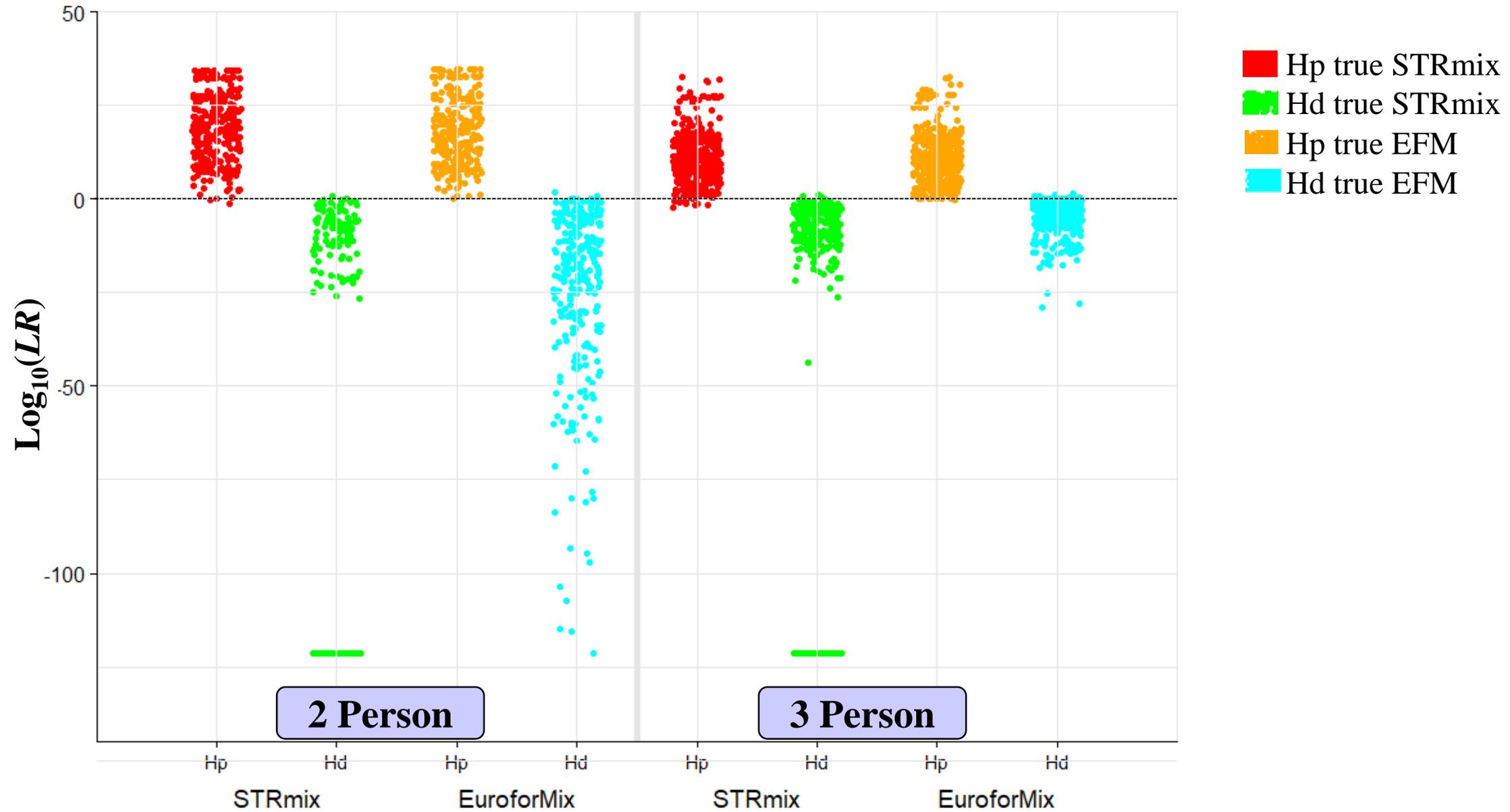
True contributors

True non-contributors ← (random person from NIST 1036)

# Analysis of 2P and 3P mixtures

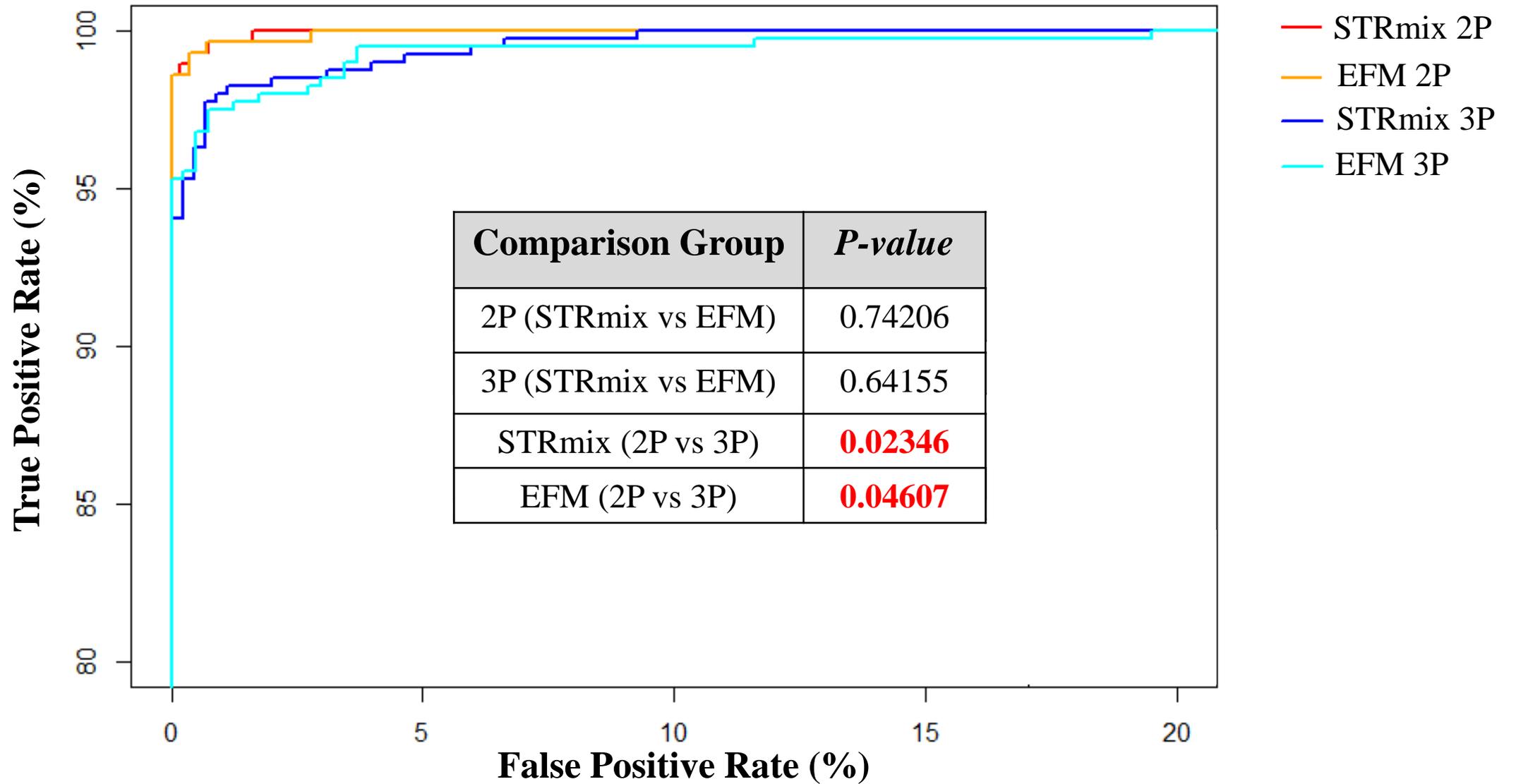
# Log<sub>10</sub>(LR) for 2P and 3P mixtures

## Log<sub>10</sub>(LR) Distribution by Software, NOC, & Propositions



# Receiver operating characteristic (ROC) plots

## ROC Plots for 2 & 3 Person Mixtures (STRmix and EFM)

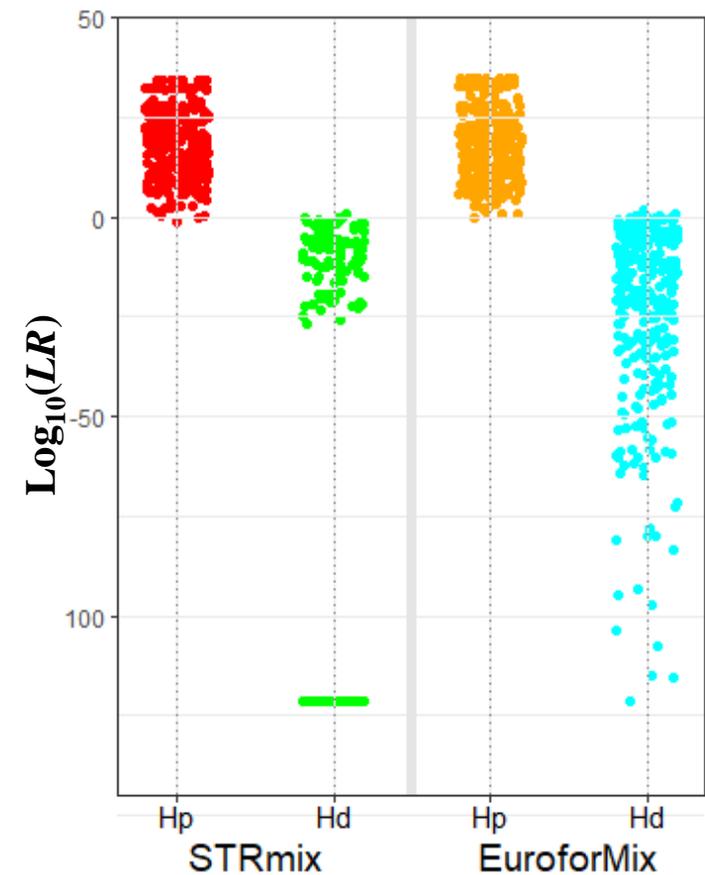


$\text{Log}_{10}(LR)$  Distribution from 2P by **software, contributor ratios**  
and **DNA treatments**

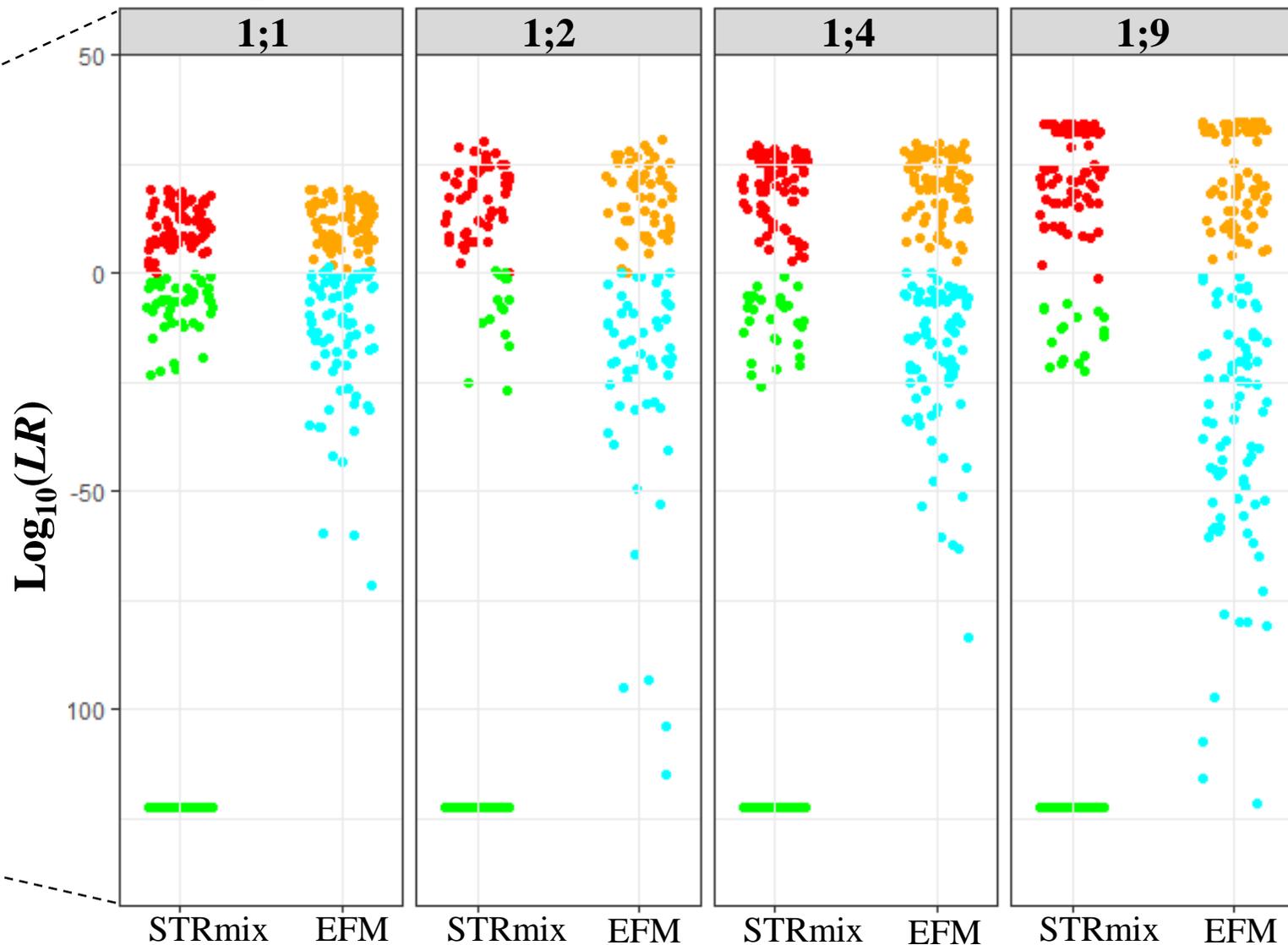
# Log<sub>10</sub>(LR) Distribution by Software & Mixture Ratios

2P

## Log<sub>10</sub>(LR) Distribution for 2P by Software & Proposition



## Log<sub>10</sub>(LR) Distribution by Software & Mixture Ratios (2P)



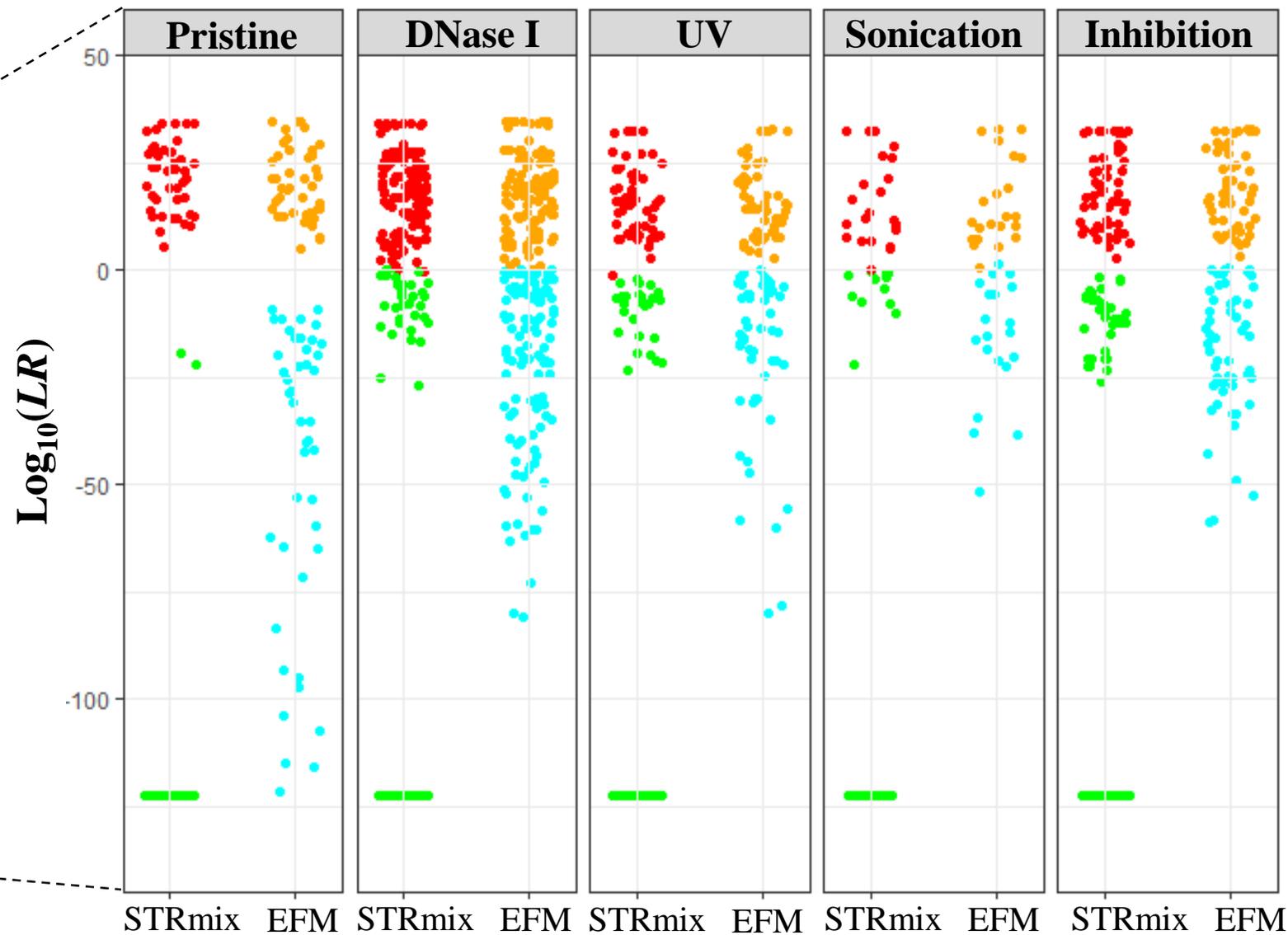
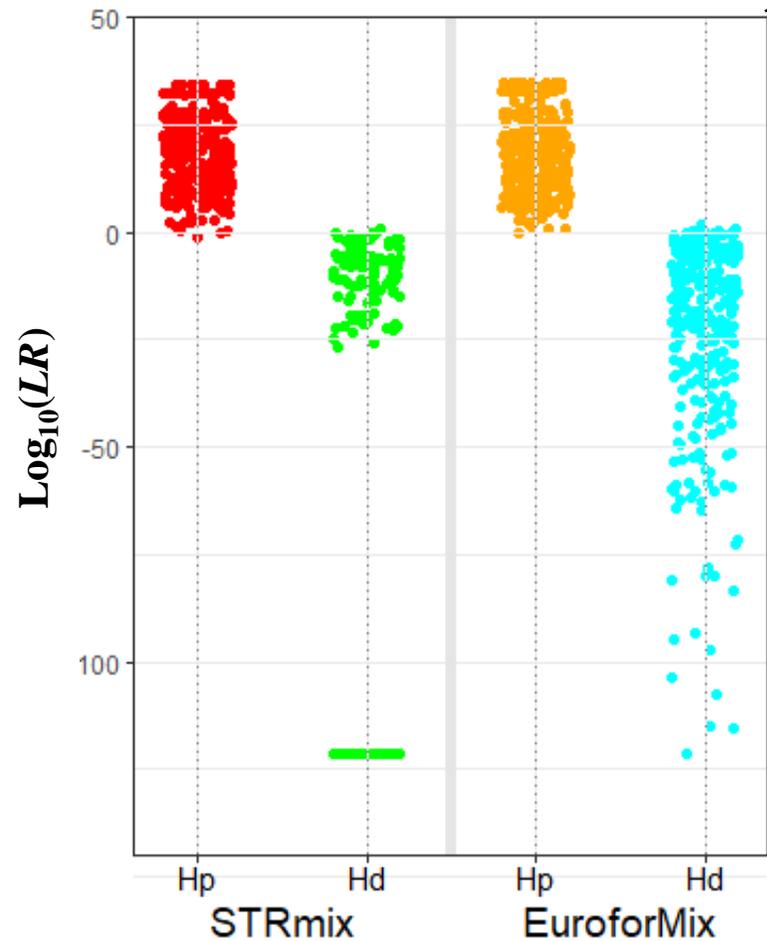
• Hp true STRmix • Hd true STRmix • Hp true EFM • Hd true EFM

# Log<sub>10</sub>(LR) Distribution by Software & Treatment

## Log<sub>10</sub>(LR) Distribution by Software & Treatment (2P)

2P

### Log<sub>10</sub>(LR) Distribution for 2P by Software & Proposition



• Hp true STRmix • Hd true STRmix • Hp true EFM • Hd true EFM

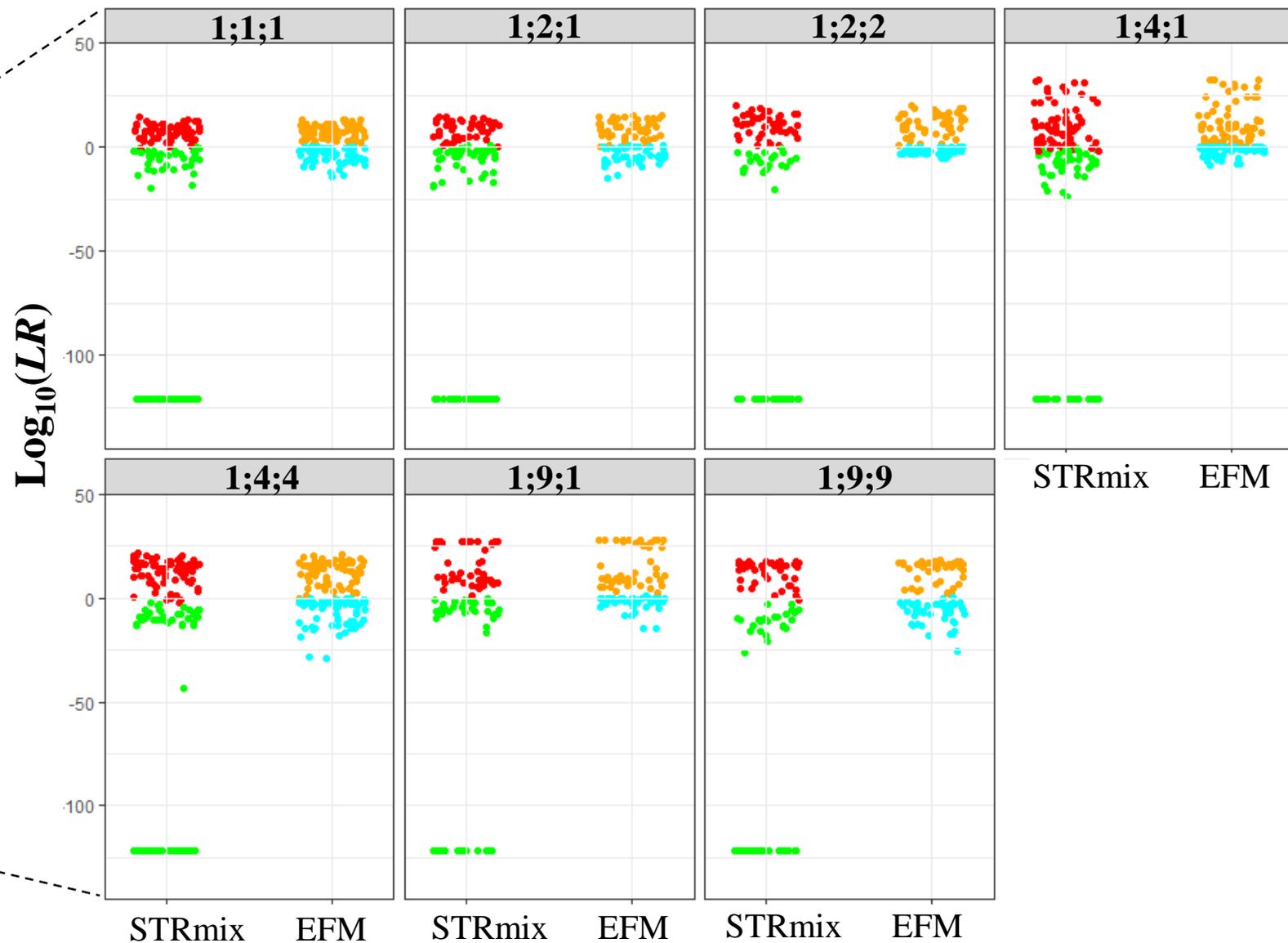
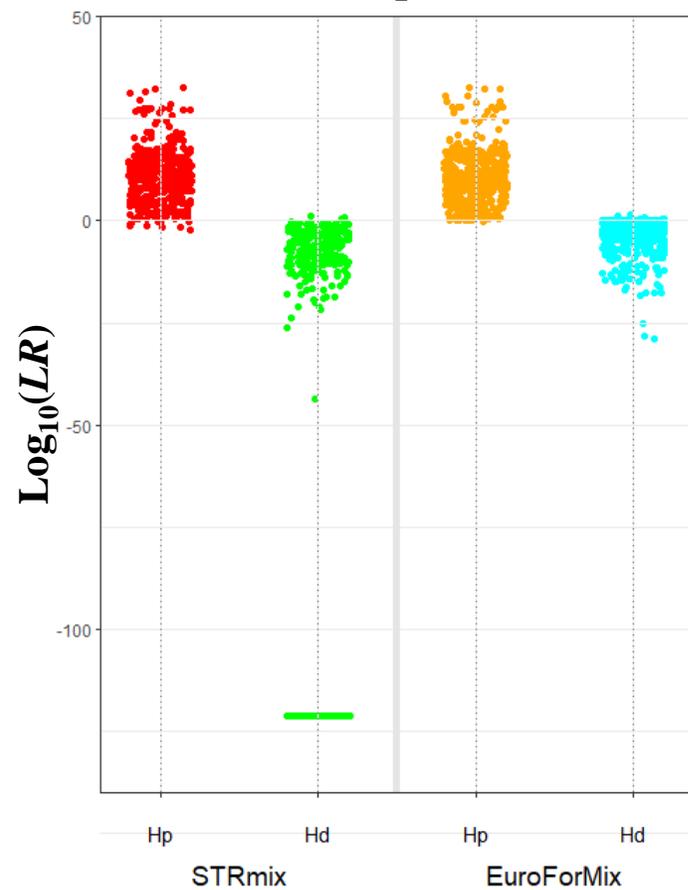
$\text{Log}_{10}(LR)$  Distribution from 3P by **software, contributor ratios**  
and **DNA treatments**

# Log<sub>10</sub>(LR) Distribution by Software & Mixture Ratios

## Log<sub>10</sub>(LR) Distribution by Software & Mixture Ratios (3P)

3P

Log<sub>10</sub>(LR) Distribution for 3P by Software & Proposition



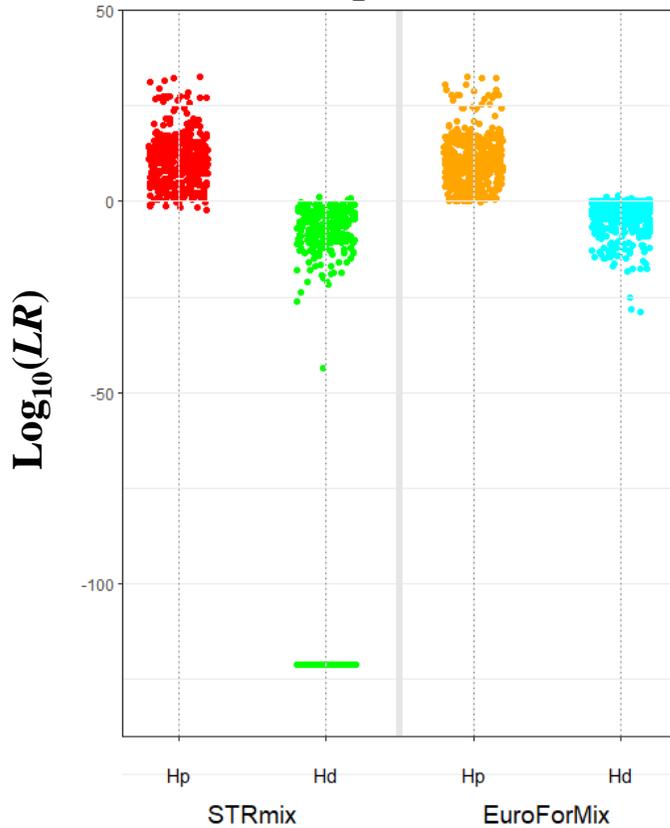
• Hp true STRmix • Hd true STRmix • Hp true EFM • Hd true EFM

# Log<sub>10</sub>(LR) Distribution by Software & Treatment

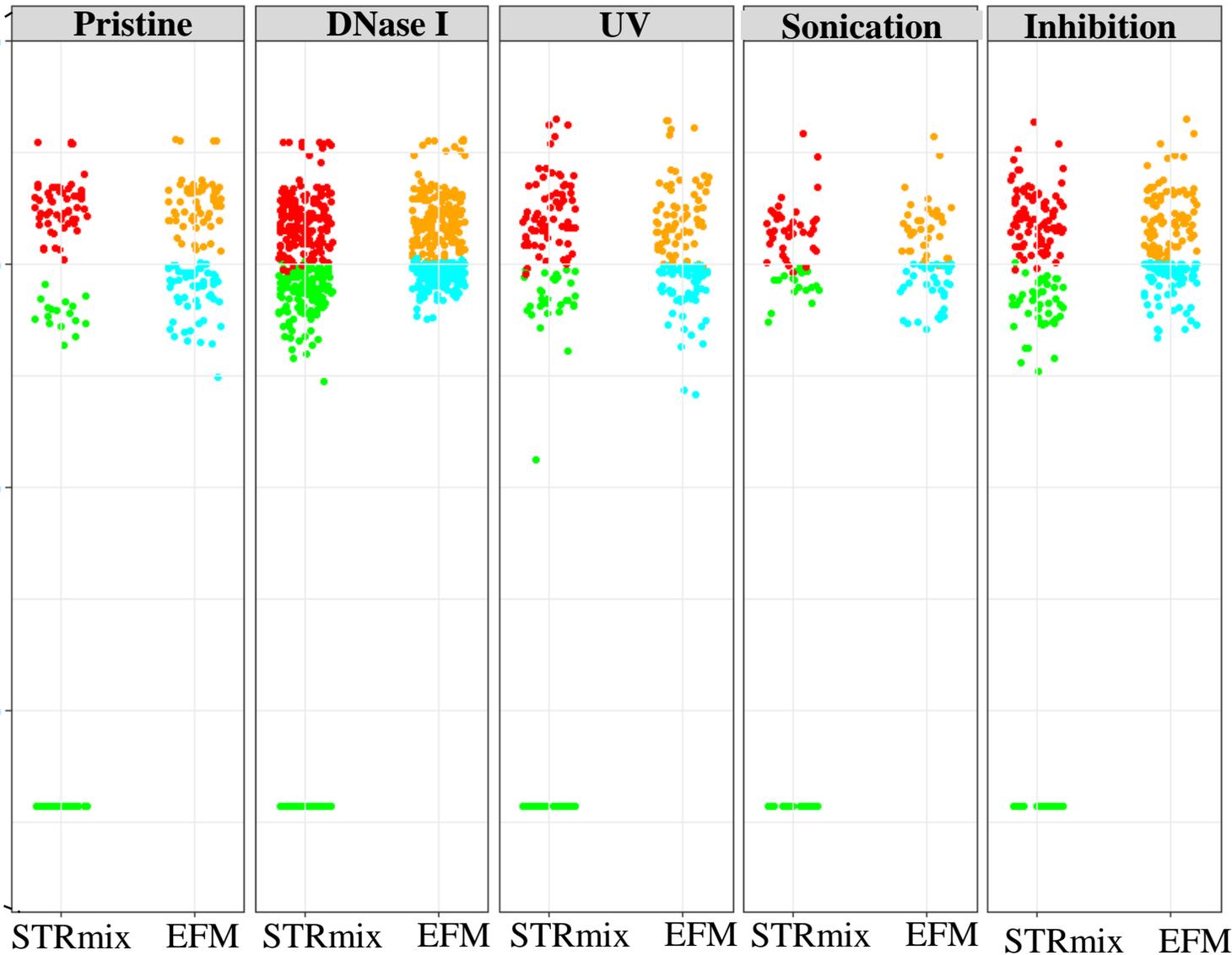
## Log<sub>10</sub>(LR) Distribution by Software & Treatment (3P)

3P

### Log<sub>10</sub>(LR) Distribution for 3P by Software & Proposition



Log<sub>10</sub>(LR)



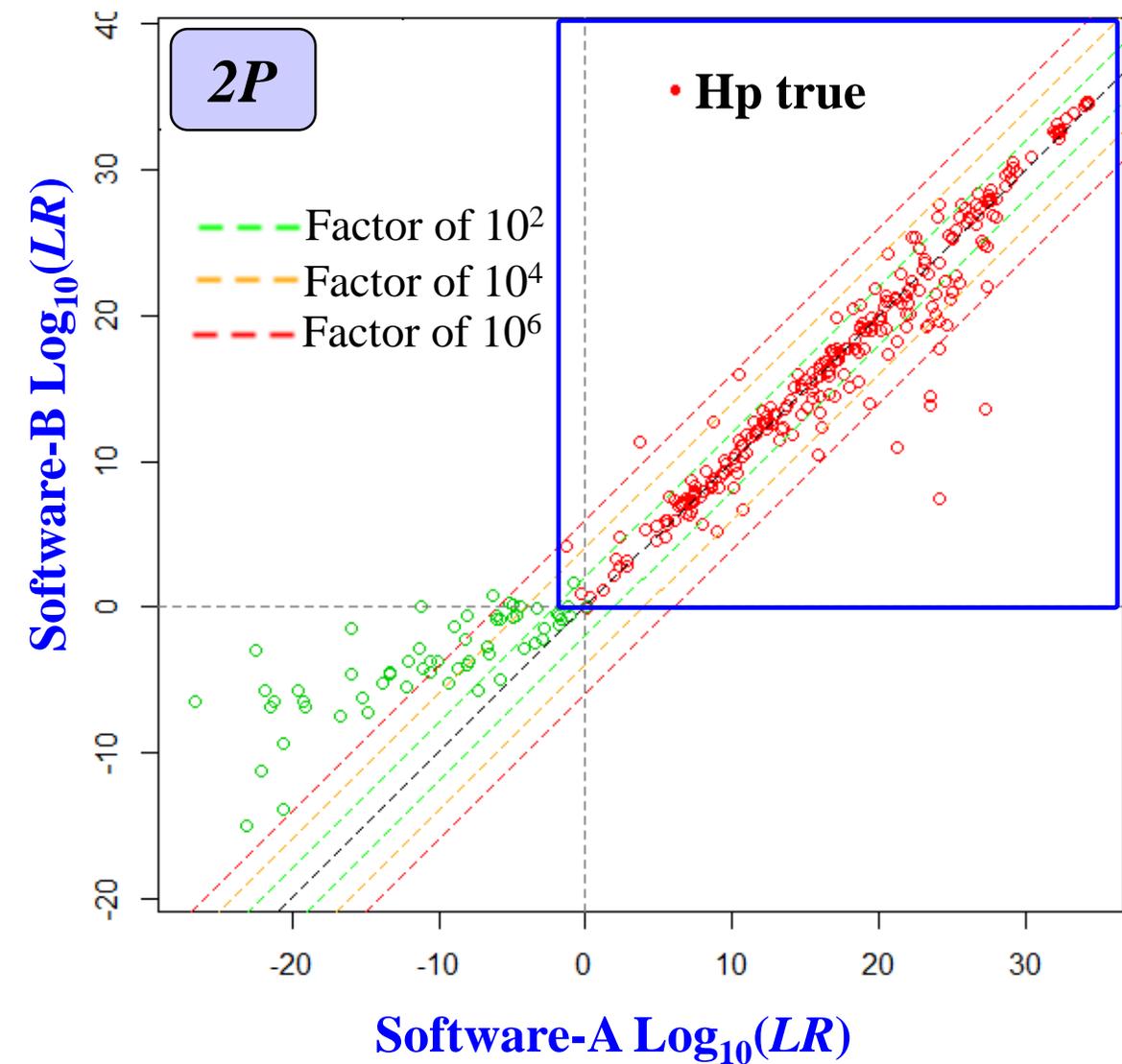
• Hp true STRmix • Hd true STRmix • Hp true EFM • Hd true EFM

**Global overall profile  $\text{Log}_{10}(LR)$  from each software**

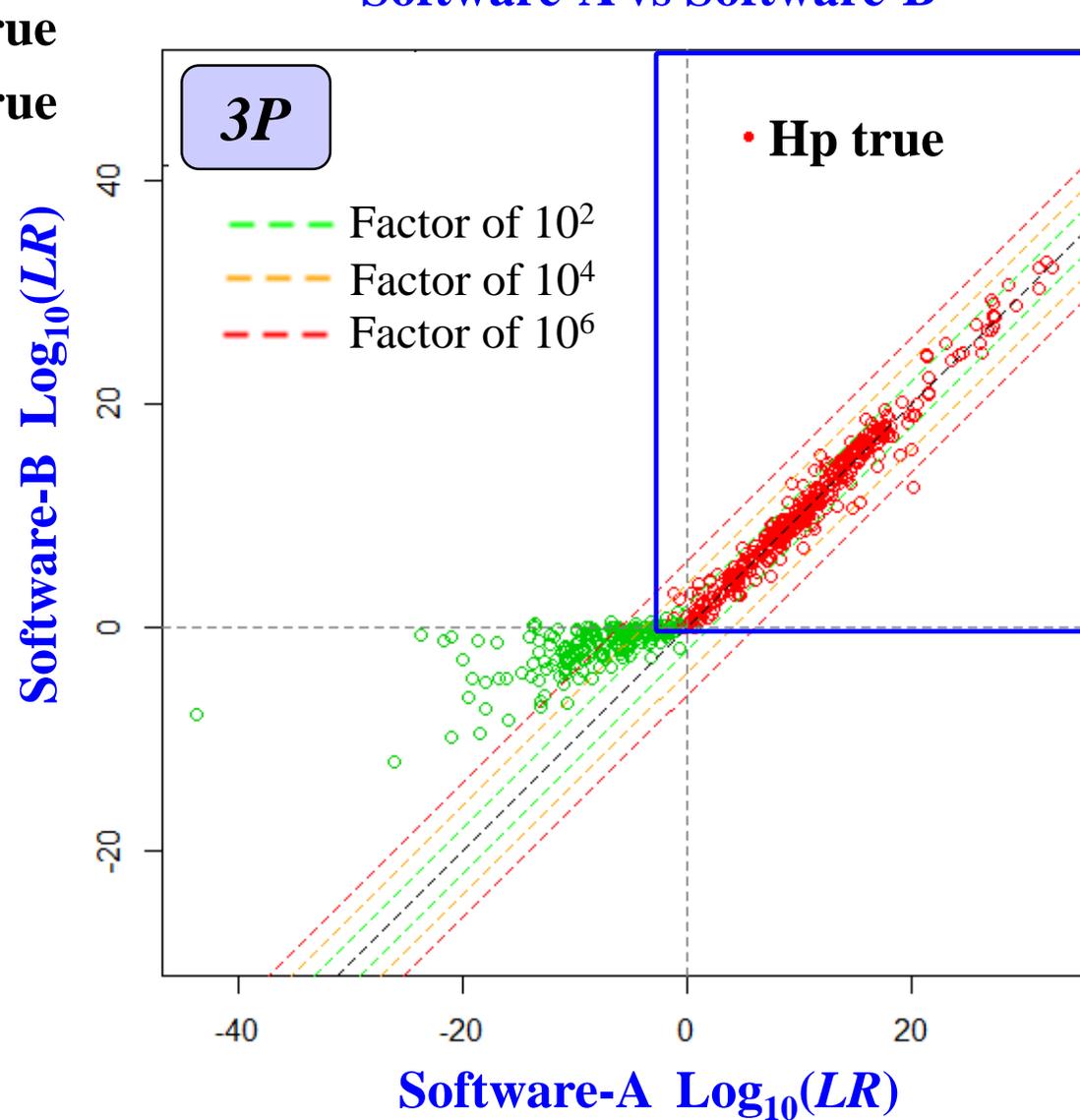
**Note for the following plots:**  $\text{Log}_{10}(LR)$  from each software is shown ‘as is’ without further designation of the sample type, ratio, treatment, or software run diagnostics (this will be addressed in future work)

# Global profile $\text{Log}_{10}(LR)$ from 2P and 3P

## Software-A vs Software-B



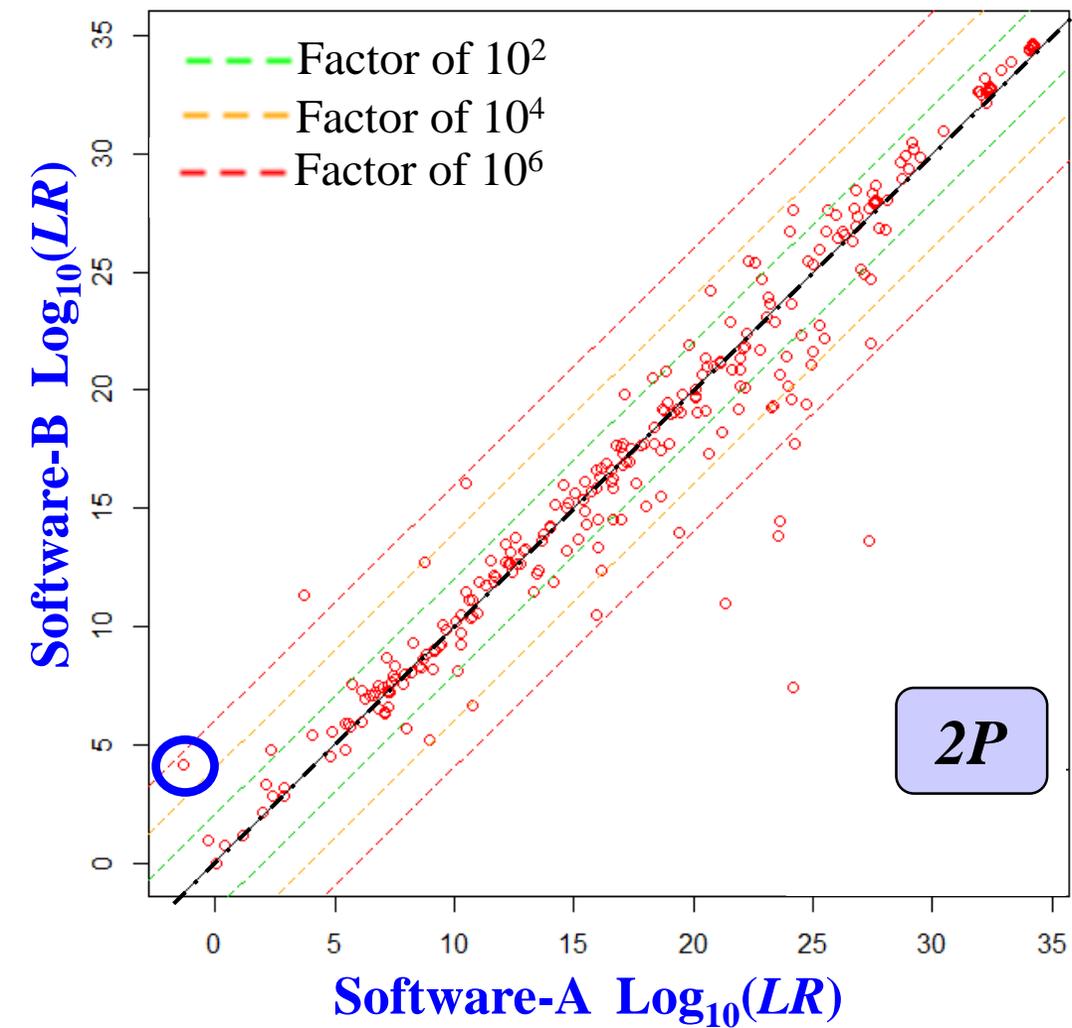
## Software-A vs Software-B



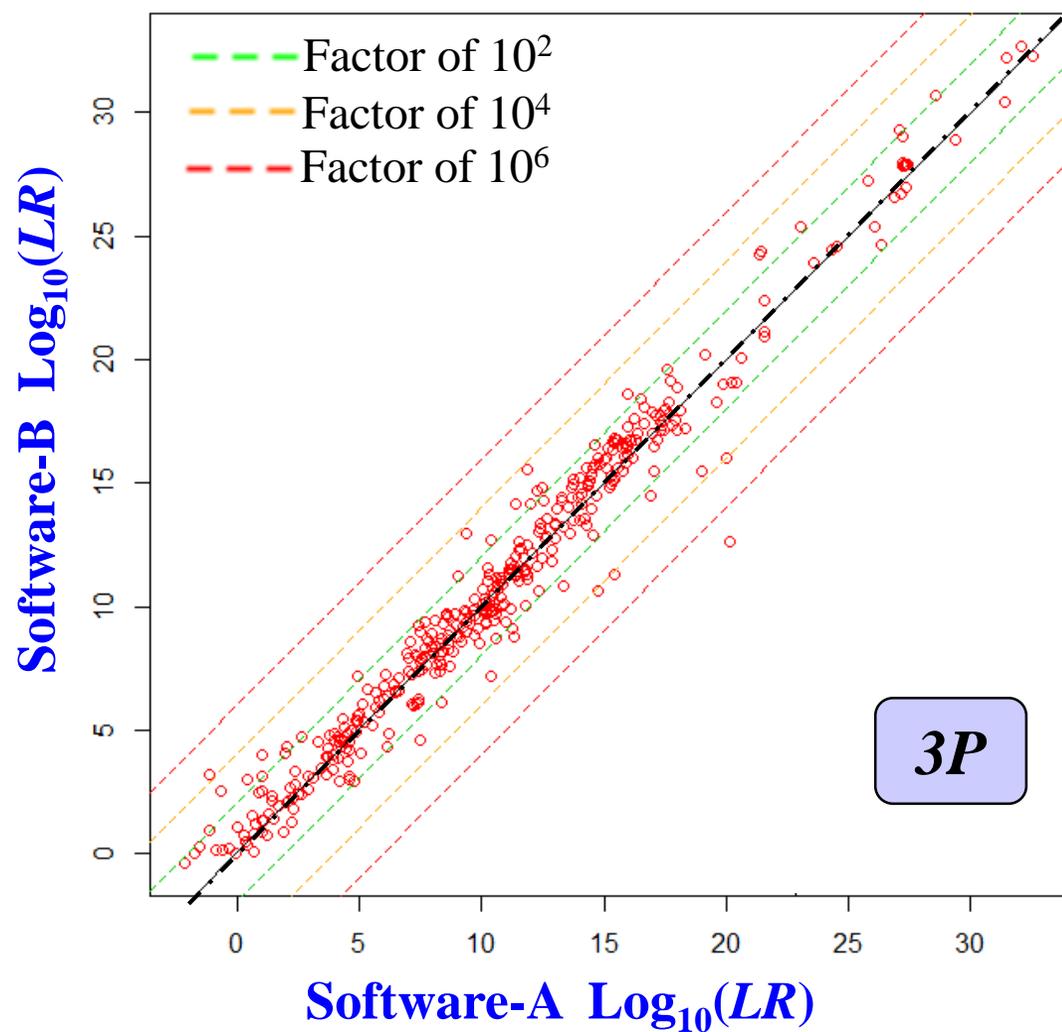
Note:  $\text{Log}_{10}(LR)$  for Hd true tests with values of  $-\infty$  from either software are not shown in these graphs

# Global profile $\text{Log}_{10}(LR)$ from 2P and 3P for **Hp true**

## Software-A vs Software-B



## Software-A vs Software-B



# Conclusions

- The publicly available PROVEDIt database is a useful resource to understand probabilistic genotyping software
- The effects of software (STRmix and EuroForMix), NOC, mixture ratios, and DNA treatments on LR assessment were examined
- As expected, both software showed high degree of discrimination between Hp TRUE and Hd TRUE distributions across different ratios and treatments for 2 and 3 contributor samples
- When it came to **sample to sample profile comparisons** the degree of agreement between the two software varied

# Future work

- Further investigation is needed to **understand the source(s) behind the LR differences** (e.g. MCMC settings, diagnostics, number of iterations, stutter models on/off, seed number)
- Analyze **additional samples** at different mixture ratios, treatments, and DNA amounts
- Explore the *4P mixtures*
- Study **deconvolution analysis** of major and minor contributors in both software
- Examine the reported LR values at a **per-locus level**

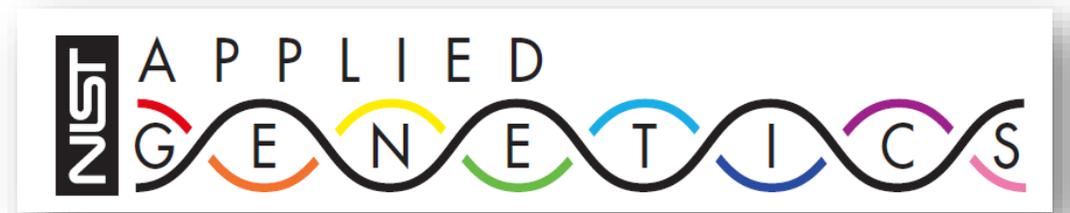
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[Contact: sarah.riman@nist.gov](mailto:sarah.riman@nist.gov)

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